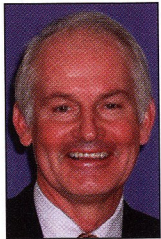


Leave Decay in My Cavity? You Must Be Kidding!



Geoffrey M. Knight, BDS, MSc, MBA, PhD

John M. McIntyre, AM, BDS, PhD

Graham G. Craig, AM, MDS, PhD

Mulyani, BDS, MDS, PhD

It is now accepted that caries infected dentin may stabilize beneath a restoration¹ and, since the late 1990s, there has been an emerging bank of evidence showing that the creation of a biological seal at the cavo margin (isolating the lesion from the overlying biofilm) reduces the viability of bacteria remaining within the lesion and arrests further caries progression.²

Despite this, it is generally accepted amongst many dentists that bacterially infected dentin is irreversibly decomposed, unable to remineralize and should be removed prior to placing a restoration. This removal of softened dentin to create a firm base for a restoration has been one of the traditional cornerstones of restorative dentistry. However, if carious dentin does not have an inhibitory effect upon invading bac-

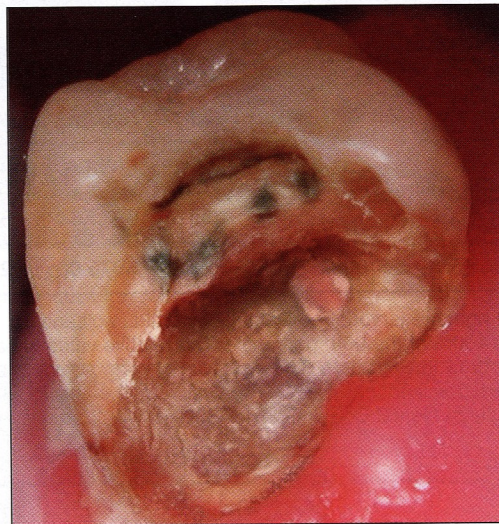


Figure 1. Arrested caries exposed beneath a lost restoration, after 18 months. Hardened tissues resisted scratching with a sharp probe.

strates the ability of a tooth to heal itself by the remineralization of carious tooth structure. Arrested caries are inevitably black in color as sulphur salts become incorporated into the remineralizing tissue. Once these lesions remineralize, they remain resistant to further caries attack unless there are dramatic changes in the oral environment. This is partly due to the remineralization process that transforms tooth dentin, carbonated apatite with a demineralization pH of around 5.5, into a complex of hydroxyl and fluorapatite⁴ (plus other ions in the oral environment) that is able to resist demineralization at a pH of around 4.5. This is a pH level that pushes the biological tolerance of many oral bacteria.

The ability of carious teeth to remineralize may be assisted by preventing biofilm formation over the lesion, without which caries are unable to progress. Both the application

...it is generally accepted amongst many dentists that bacterially infected dentin is irreversibly decomposed, unable to remineralize and should be removed prior to placing a restoration.

teria, it infers that the dentin pulp complex is the only vital tissue in the body without a front line physiological response to bacterial invasion. What are the evolutionary parameters that maintain a single point of tissue vulnerability in the body against infection?

A recent study³ has shown that demineralized dentin was more effective in reducing the viability of *streptococcus mutans* than dentin treated with an antibacterial substance (silver fluoride [AgF] and potassium iodide [KI]). This suggests there are sub-

stances that may be released by carious dentin that inhibit bacteria viability. When AgF and KI are applied to the surface of demineralized dentin there is a further substantial reduction in bacterial viability³ and suggests that the application of AgF and KI works synergistically with demineralized dentin in further reducing the reproductive potential of the bacteria.

ARRESTED ROOT CARIES

The presence of arrested root caries demon-

of ozone⁵ and AgF/KI⁶ to dentin prevents biofilm formation. This is analogous to dressing a soft tissue wound with materials such as silver ointment or iodide to assist healing.

In caries affected dentin some demineralization occurs, but the collagen matrix remains intact enabling reconstitution of a hydroxyl fluorapatite dentin.

Strontium and fluoride ions from glass ionomer (GI) cement restorations have been detected in infected dentin consistent with remineralization.⁷ The nature and composition of the remineralized tissues will depend upon the ions present and the extent of degradation of the supporting collagen matrix.

The hardening of carious dentin may be compared to skin scar tissue formation.

continued on page 132

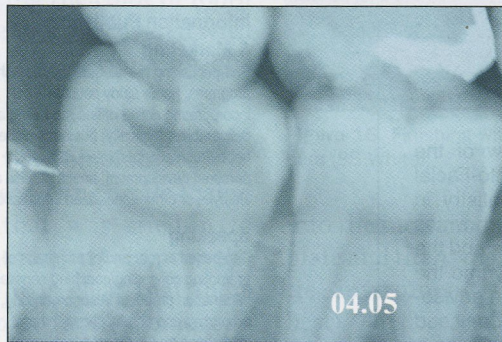


Figure 2. A patient presented with a large asymptomatic lesion that showed extensive caries radiographically.



Figure 3. The dentin was treated with HealOzone (KaVo), and a self-curing glass ionomer (GI) cement restoration was then placed over the lesion.



Figure 4. Twelve months after placement, one can see that there was a significant increase in radiopacity within the carious dentin.

Leave Decay in My Cavity?...

continued from page 130

Figure 1 shows arrested caries after 18 months beneath a lost GI cement restoration. There is no black staining since the lesion was isolated from the oral environment. The surface of the lesion has a cracked appearance due to the dehydration of the hydrated carious dentin during the hardening process. The surface of the lesion is obviously not remineralized dentin, but a tissue hard enough to resist marking with a sharp probe.



Figure 5. A carious lesion was evident on the occlusal surface of a lower molar.



Figure 6. Enamel was removed to gain access to the caries and a moat was prepared in sound dentin at the dentin-enamel junction with a No. 3 round slow speed bur.



Figure 8. A 3cm square piece of thin flexible plastic was placed over the preparation. The patient held this in centric occlusion for 4 minutes until the GI cement had cured.



INFORMED CONSENT IS IMPORTANT

Intentionally leaving caries under a restoration may have the potential to lead to legal problems if a patient is unaware of the nature of procedure. This is especially true if another practitioner should have to radiograph the restoration and be unaware of this treatment protocol. Dentists who carry out remineralization procedures are well advised to provide their patients with written explanatory notes about the procedure and the benefits that can be achieved.

CONCLUSION

Restoration of teeth by amputation is a caries management model that often leads to the ongoing iatrogenic destruction of the dentition. The pharmacological management of caries is a conservative alternative that enables the remineralization of caries infected teeth to form a decay resistant layer at the base of a restoration.

AgF has been used to arrest caries, primarily in deciduous teeth since the early 1970s. After application, free silver ions react with oral sulphides to form silver sulphide, staining the teeth black. The application of KI

composite resins which offers no such protection.⁹

The following clinical case shows increased radiopacity that occurred under a self-curing GI cement restoration treated with ozone.

A patient presented with a large carious lesion on a lower second molar (Figure 2). Although the lesion was asymptomatic, a periapical radiograph showed extensive caries that

Dentists who carry out remineralization procedures are well advised to provide their patients with written explanatory notes about the procedure and the benefits....

may well have resulted in a pulp exposure during cavity preparation.

Overlying caries were removed and the lesion was etched with phosphoric acid for 5 seconds, then washed and dried with oil-free air. The residual softened caries were treated with ozone for 40 seconds using a HealOzone (KaVo) unit. Following this the cavity was restored with a self-curing GI cement (Fuji Triage [GC

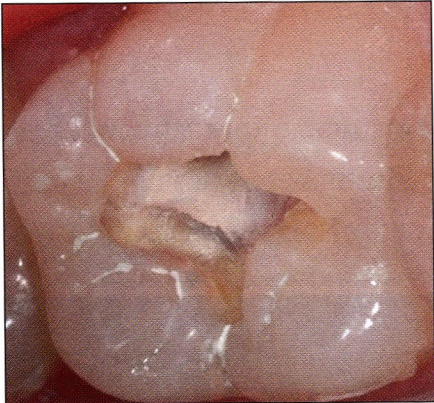


Figure 7. After etching, silver fluoride was applied to the preparation, followed by potassium iodide. The cavity was then washed and dried with oil-free air. A creamy-white precipitate formed on the remaining carious dentin.

America)]. Extensive radiolucency below the restoration is apparent, immediately after restoration placement (Figure 3). However, 12 months later, when the patient was recalled to have a composite resin (Ice [Southern Dental Industries]) placed over the GI, a further radiograph shows a marked improvement in radiopacity below the GI base (Figure 4).

Case Report 2: Glass Ionomer Used With Silver Fluoride and Potassium Iodide

The following case report describes a pharmacological approach to managing a carious lesion using AgF/KI and GI cement:

Figure 9. After the GI was cured, minimal contouring was required to complete the restoration.

The patient presented with occlusal caries on a lower molar in Figure 5.

Enamel was removed to gain access to the lesion. A moat was then prepared in sound dentin with a No. 3 slow speed round bur, at the dentin-enamel junction (DEJ) around the perimeter of the caries (Figure 6). Next, the preparation was etched with 37% phosphoric acid for 5 seconds, washed with water, and dried with oil-free air. AgF was then applied to the preparation, followed by KI until the precipitation turned from cloudy white to clear. The cavity was washed and dried again with oil-free air (Figure 7).

After isolating the preparation from moisture with strategically placed cotton rolls, a self-curing GI cement (Riva [Southern Dental Industries]) was placed into the preparation to slightly overfill the preparation. AgF/KI enhances the bond strength between self-curing GI and dentin.¹⁰ A 3 cm square piece of a plastic freezer bag was placed over the GI and the patient asked to close in into centric occlusion for 4 minutes until the GI had cured (Figure 8). After curing, minimal contouring was required to complete the restoration (Figure 9).

immediately after AgF application forms silver iodide. This is a low-solubility creamy-white precipitate with significant antibacterial properties that inhibits silver staining.♦

References

1. Mertz-Fairhurst EJ, Curtis JW Jr, Ertle JW, et al. Ultraconservative and cariostatic sealed restorations: results at year 10. *J Am Dent Assoc.* 1998;129:55-66.
2. Kidd EA. How "clean" must a cavity be before restoration? *Caries Res.* 2004;38:305-313.
3. Knight GM, McIntyre JM, Craig GG, et al. Differences between normal and demineralized dentine pretreated with silver fluoride and potassium iodide after an in vitro challenge by *Streptococcus mutans*. *Aust Dent J.* 2007;52:16-21.
4. Chow LC, Vogel GL. Enhancing remineralization. *Oper Dent.* 2001;26(suppl 6):27-38.
5. Knight GM, McIntyre JM, Craig GG, et al. The inability of *Streptococcus mutans* and *Lactobacillus acidophilus* to form a biofilm in vitro on dentine pretreated with ozone. *Aust Dent J.* 2008;53:349-353.
6. Knight GM, McIntyre JM, Craig GG, et al. Inability to form a biofilm of *Streptococcus mutans* on silver fluoride- and potassium iodide-treated demineralized dentin. *Quintessence Int.* 2009;40:155-161.
7. Ngo HC, Mount G, McIntyre J, et al. Chemical exchange between glass-ionomer restorations and residual carious dentine in permanent molars: an in vivo study. *J Dent.* 2006;34:608-613.
8. Knight GM, McIntyre JM, Craig GG, et al. Electron probe microanalysis of ion exchange of selected elements between dentine and adhesive restorative materials. *Aust Dent J.* 2007;52:128-132.
9. Knight GM, McIntyre JM, Craig GG, et al. An in vitro investigation of marginal dentine caries abutting composite resin and glass ionomer cement restorations. *Aust Dent J.* 2007;52:187-192.
10. Knight GM, McIntyre JM, Mulyani. The effect of silver fluoride and potassium iodide on the bond strength of auto cure glass ionomer cement to dentine. *Aust Dent J.* 2006;51:42-45.

MINIMALLY INVASIVE DENTISTRY

Dr. Knight is a general dentist and internationally noted dental speaker from Melbourne, Australia with special interests in minimal intervention aesthetic dentistry. He has pioneered techniques for the pharmacological management of dental caries and periodontal disease. He has introduced a number of innovative clinical techniques for aesthetic enhancement with minimal tooth preparation and he is named on several dental patents. Apart from his broad clinical base, he has been State President of his Dental Association and has extensive political and economic experience within the profession. He is well focused on the many problems of dentistry and the solutions needed to survive today's rapidly changing environment. He has been published in *Quintessence International*, *Australian Dental Journal*, and *Journal of Periodontal Research*. He has produced a series of clinical videos and written numerous articles on aesthetic and adhesive dentistry that have been translated and published in a number of languages. He can be reached via e-mail at geoffbds@dentalk.com.au.

Disclosure: Drs. Knight and Craig are named on a process patent associated with the use of silver fluoride and potassium iodide. Dr. Knight was associated with the development of Fuji Triage and has a financial interest in this product.

nesia. She received her Diploma in Clinical Dentistry, her Master of Dental Surgery (Pedodontics), and Doctor of Philosophy (Community Dentistry) from the University of Adelaide in Australia. Currently, Dr. Mulyani works mainly in dental material research while providing supervision on research methodology for post graduate students in

Dental School, The University of Adelaide. Her research interests include cariology, glass ionomer cements, erosion, and minimally invasive restorative dentistry. She was a lecturer in the Faculty of Dentistry, University of North Sumatera in Indonesia for 20 years. She has published handbooks and manuals for the students to be used in her teaching.

She also published papers in dental journals and presented papers and posters in Indonesian Dental conferences and IADR conferences. She can be reached via e-mail at mulyani.dalidjan@adelaide.edu.au.

Disclosure: Dr. Mulyani reports no conflicts of interest.

Dr. McIntyre received his primary dental degree from University of Queensland in 1960 and his PhD in Microbiology and Immunology at Adelaide University in 1970, both in Australia. He is the former Dean of Adelaide Dental School and Chairman of the department of Dentistry. Currently retired, Dr. McIntyre is a part-time Visiting Research Fellow at the University, and supervising post-graduate research projects at all levels. His current research involvement includes cariology, dental erosion, glass ionomer cements; fluoride action and vehicles for use in developing countries, minimally invasive methods in restorative dentistry. He can be reached at john.mcintyre@adelaide.edu.au.

Disclosure: Dr. McIntyre reports no conflicts of interest.

Dr. Craig was formerly associate professor in Preventive Dentistry at the University of Sydney and, after that, director of the Dental Health Foundation in Australia. He has had extensive involvement in the promotion of water fluoridation and in programs designed to improve the dental health of the community including various aspects of minimal intervention dentistry. He has published in numerous scientific journals and has presented more than 200 post-graduate courses and lectures. He can be reached at dentaloutlook@bigpond.com.

Disclosure: Drs. Knight and Craig are named on a process patent associated with the use of silver fluoride and potassium iodide.

Dr. Mulyani received her Bachelor of Dental Surgery from the University of North Sumatera in Indo-